

## Supplementary Materials for

### **A comprehensive portrait of the venom of the giant red bull ant, *Myrmecia gulosa*, reveals a hyperdiverse hymenopteran toxin gene family**

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#### **Other Supplementary Material for this manuscript includes the following:**

(available at [advances.sciencemag.org/cgi/content/full/4/9/eaau4640/DC1](https://advances.sciencemag.org/cgi/content/full/4/9/eaau4640/DC1))

Movie S1 (.mov format). Collection of venom from *M. gulosa*.

## Supplementary Materials and Methods

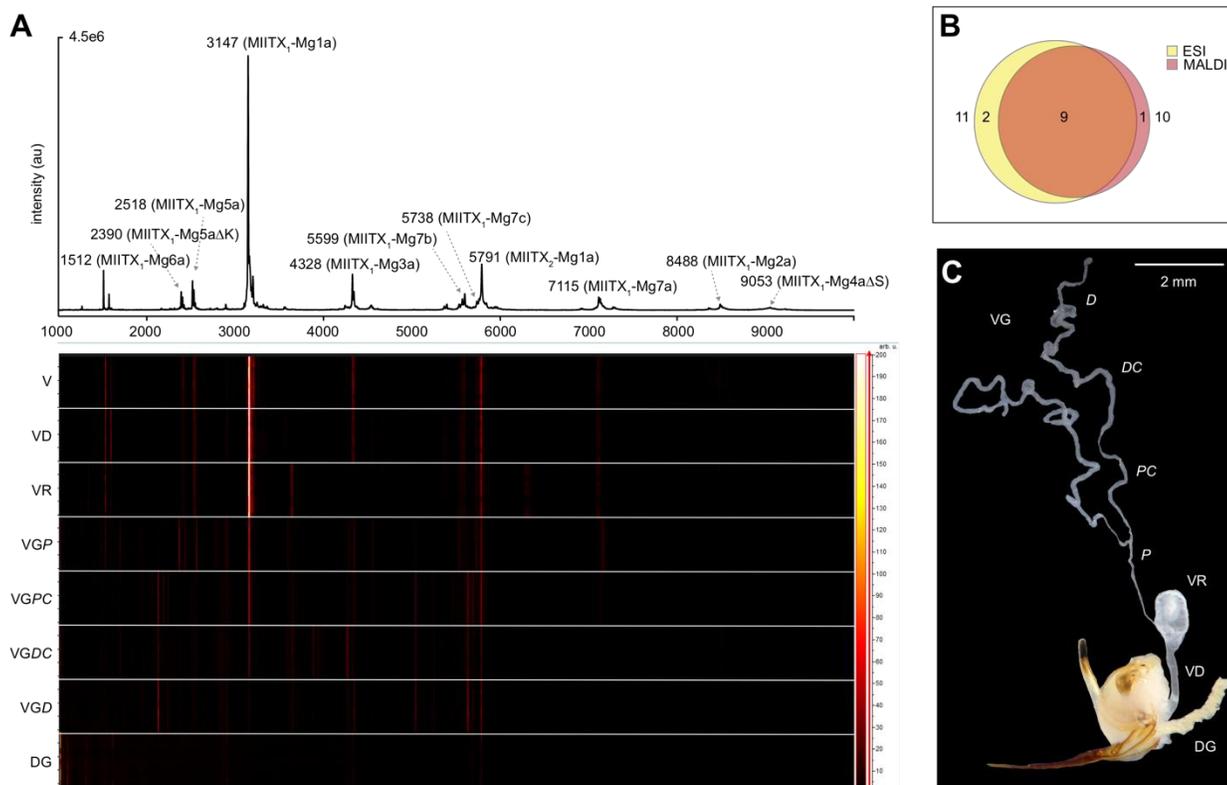
### Antimicrobial and cytotoxicity assays

Antimicrobial and cytotoxicity assays were performed by CO-ADD (The Community for Antimicrobial Drug Discovery). Briefly, growth inhibition of all bacterial strains was determined by measuring absorbance at 600 nm, growth inhibition of *C. albicans* was determined by measuring absorbance at 530 nm, and growth inhibition of *C. neoformans* was determined by measuring the difference in absorbance between 600 and 570 nm, after the addition of resazurin (0.001% final concentration) and incubation at 35°C for an additional 2 h. Growth inhibition of HEK293 cells was determined by measuring fluorescence (excitation 530/10 nm, emission 590/10 nm) after the addition of resazurin (25 µg/mL final concentration) and incubation at 37°C and 5% CO<sub>2</sub>, for an additional 3 h. Percentage growth inhibition was calculated using negative controls (media only) and positive controls (no peptide). For the haemolytic activity assay, human whole blood was washed three times with 3 volumes of 0.9% NaCl and then resuspended in 0.9% NaCl to a concentration of  $0.5 \times 10^8$  cells/mL. Cells were incubated for 1 h at 37°C with or without the peptide. After incubation, plates were centrifuged at 1000 g for 10 min to pellet cells and debris and haemolysis determined by measuring the supernatant absorbance at 405 nm. Minimum inhibitory concentration (MIC), CC<sub>50</sub> (concentration at 50% cytotoxicity) and HC<sub>50</sub> (concentration at 50% haemolytic activity) values were calculated by curve fitting the inhibition values versus log(concentration) using a sigmoidal dose-response function (variable slope), in Pipeline Pilot's dose-response component.

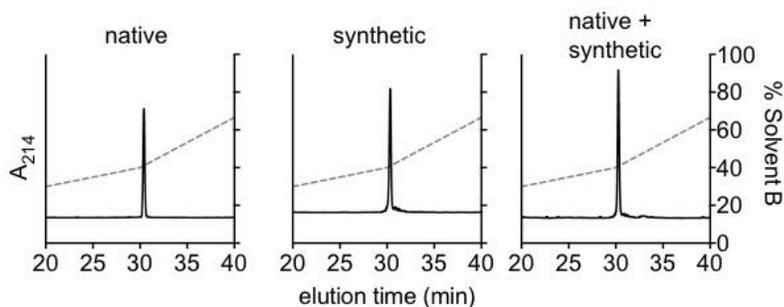
## Results

### Antimicrobial activity of MIITX<sub>1</sub>-Mg1a

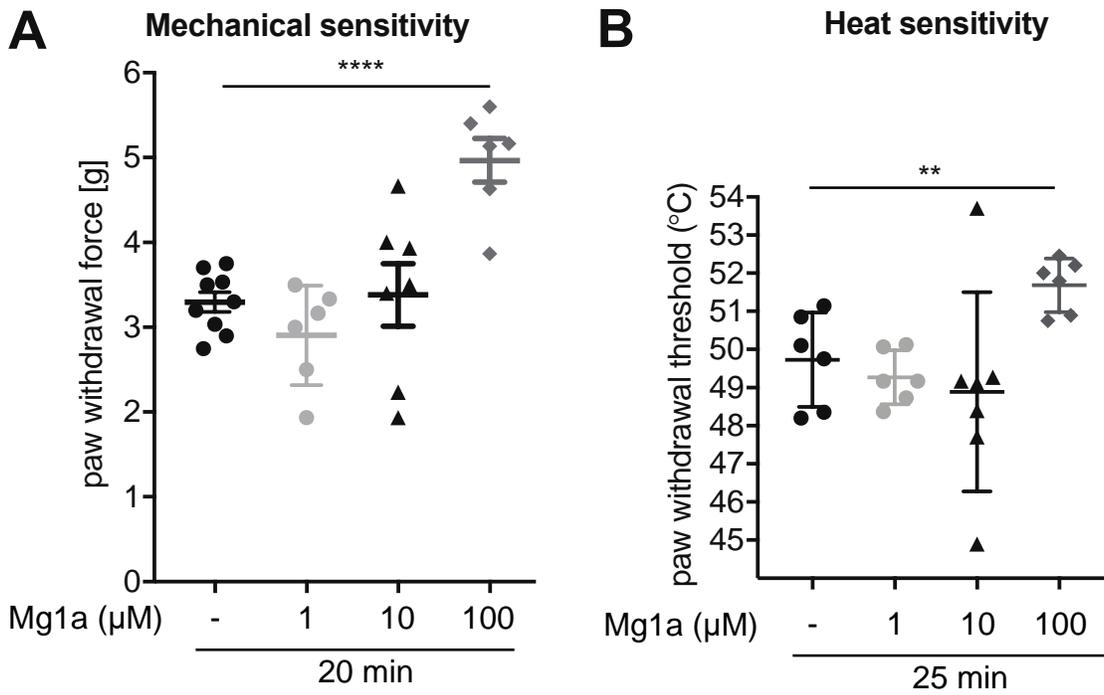
Several previously described ant venom peptides have been shown to have antimicrobial activity (20), and therefore we examined the potential utility of MIITX<sub>1</sub>-Mg1a as an antimicrobial. MIITX<sub>1</sub>-Mg1a was tested on a range of available microbial strains (**Table S2**). The peptide caused complete growth inhibition of the fungus *Cryptococcus neoformans var. grubii*, with an MIC of 2.5 µM. At the highest concentration tested (10.2 µM), it also partially inhibited growth of the bacteria *Staphylococcus aureus* and *Acinetobacter baumannii*, with 84 and 32% inhibition, respectively, but had no effect on the bacteria *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, or the fungus *Candida albicans*. At the highest concentration tested (10.2 µM), MIITX<sub>1</sub>-Mg1a was not haemolytic (to human red blood cells), but it did display partial toxicity (49.1% growth inhibition) of HEK293 cells.



**Fig. S1. MALDI-MS analysis of *M. gulosa* venom apparatus.** (A) MALDI spectrum, generated in linear positive mode, of *M. gulosa* venom. Peaks corresponding to peptides identified as venom components by combined ESI-MS/transcriptomics are labelled. Theoretical  $MH_{av}^{+1}$  for each peptide are as follows: MIITX<sub>1</sub>-Mg6a, 1511.9; MIITX<sub>1</sub>-Mg5aΔK, 2390.9; MIITX<sub>1</sub>-Mg5a, 2519.1; MIITX<sub>1</sub>-Mg1a, 3147.8; MIITX<sub>1</sub>-Mg3a, 4329.0; MIITX<sub>1</sub>-Mg7b, 5600.4; MIITX<sub>1</sub>-Mg7c, 5740.6; MIITX<sub>2</sub>-Mg1a, 5792.3; MIITX<sub>1</sub>-Mg7a, 7116.3; MIITX<sub>1</sub>-Mg2a, 8489.4; MIITX<sub>1</sub>-Mg4aΔS, 9054.1. Gel view (prepared in ClinProTools) illustrates variation across 10 individual shots of samples prepared from venom (V), venom duct (VD), venom reservoir (VR), Dufour's gland (DG) and 4 sections of the venom glands: proximal (VGP), proximal central (VGPC), distal central (VGDC) and distal (VGD). (B) Venn diagram illustrating overlap in peptides identified using different MS techniques. (C) Venom apparatus of *M. gulosa*, labelled as above.



**Fig. S2. Coelution of native (purified from venom) and synthetic MIITX<sub>1</sub>-Mg1a.** RP-HPLC was performed using a Phenomenex Gemini NX-C<sub>18</sub> column (250 x 4.6 mm, 3 μm particle size, 110 Å pore size) with a gradient of 15–45% solvent B (90% ACN, 0.05% TFA) over 30 min.



**Fig. S3. MIITX<sub>1</sub>-Mg1a-induced changes in paw withdrawal.** (A) At a concentration of 100  $\mu\text{M}$ , MIITX<sub>1</sub>-Mg1a had caused mechanical sensitivity (at 20 min post-injection),  $n = 3-9$  per group. (B) MIITX<sub>1</sub>-Mg1a (100  $\mu\text{M}$ ) caused heat sensitivity (at 25 min post-injection).  $n = 6-7$  per group. Data are expressed as mean  $\pm$  SEM. Statistical significance compared to vehicle controls was determined using one-way ANOVA with Dunnett's multiple comparison test. \*\*  $p < 0.01$ , \*\*\*\*  $p < 0.0001$ .

**Table S1. Venom- or toxin-associated annotation of nonvenom component transcripts.**

<b>UniProt Entry</b>	<b>Protein name</b>	<b>Organism</b>	<b>TPM</b>
<b>A7X3V4</b>	Kunitz-type serine protease inhibitor kunitoxin-Tell	<i>Telescopus dhara</i>	269.55
<b>B2D0J5</b>	Venom carboxylesterase-6	<i>Apis mellifera</i>	203.1
<b>C9WMM5</b>	Venom serine carboxypeptidase	<i>Apis mellifera</i>	152.1
<b>Q58L94</b>	Venom prothrombin activator notecarin-D2	<i>Notechis scutatus scutatus</i>	85.64
<b>Q58L94</b>	Venom prothrombin activator notecarin-D2	<i>Notechis scutatus scutatus</i>	70.75
<b>B2D0J5</b>	Venom carboxylesterase-6	<i>Apis mellifera</i>	54.33
<b>B5U2W0</b>	Venom serine protease Bi-V	<i>Bombus ignitus</i>	35.14
<b>Q8T0W5</b>	PIMHY Cysteine-rich venom protein 1	<i>Pimpla hypochondriaca</i>	29.79
<b>P35779</b>	Venom allergen 3	<i>Solenopsis richteri</i>	27.35
<b>A7X3V4</b>	Kunitz-type serine protease inhibitor kunitoxin-Tell	<i>Telescopus dhara</i>	19.98
<b>Q8MQS8</b>	Venom serine protease 34	<i>Apis mellifera</i>	19.82
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	18.3
<b>C9WMM5</b>	Venom serine carboxypeptidase	<i>Apis mellifera</i>	15.79
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	13.95
<b>Q8MQS8</b>	Venom serine protease 34	<i>Apis mellifera</i>	13.06
<b>Q8JI39</b>	Cysteine-rich venom protein triflin	<i>Protobothrops flavoviridis</i>	10.95
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	6.8
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	6.71
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	6.63
<b>P0DM55</b>	Venom peptide SjAPI	<i>Scorpiops jendeki</i>	5.64
<b>Q8MQS8</b>	Venom serine protease 34	<i>Apis mellifera</i>	5.05
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	4.94
<b>P505486</b>	Conopressin	<i>Conus geographus</i>	3.95
<b>B5AJT4</b>	Venom metalloproteinase 3	<i>Eulophus pennicornis</i>	3.35
<b>P35775</b>	Venom allergen 2	<i>Solenopsis invicta</i>	1.61

**Table S2. Assessment of antimicrobial, cytotoxic and hemolytic activity of MIITX<sub>1</sub>-Mg1a.**

<b>Bacterium</b>	<b>Strain</b>	<b>Organism</b>	<b>Max response ± SEM</b>	<b>MIC (μM)</b>
<i>Staphylococcus aureus</i>	ATCC 43300	Bacterium	84 ± 0	10.2
<i>Escherichia coli</i>	ATCC 25922	Bacterium	-1 ± 2	>10.2
<i>Klebsiella pneumoniae</i>	ATCC 700603	Bacterium	16 ± 1	>10.2
<i>Acinetobacter baumannii</i>	ATCC 19606	Bacterium	32 ± 3	>10.2
<i>Pseudomonas aeruginosa</i>	ATCC 27853	Bacterium	4 ± 4	>10.2
<i>Candida albicans</i>	ATCC 90028	Fungus	8 ± 4	>10.2
<i>Cryptococcus neoformans var. grubii</i>	H99; ATCC 208821	Fungus	108 ± 6	2.5
<b>Human embryonic kidney cells</b>	ATCC CRL-1573	Human	49 ± 7	8.8 (CC <sub>50</sub> )
<b>Human red blood cells</b>		Human	10 ± 1	>10.2 (HC <sub>50</sub> )

MIC, minimum inhibitory concentration; CC<sub>50</sub>, concentration at 50% cytotoxic activity; HC<sub>50</sub>, concentration at 50% haemolytic activity.